

# Gut

---

## Leading article

---

### The molecular revolution – coming your way soon

The face of molecular biology is rapidly changing from that of an exclusive scientific club using complex, expensive high technology to a simpler, cheaper discipline which can be routinely employed by the gastroenterologist. Why is this and why now? There are two main reasons. Firstly, molecular biology has begun to deliver its promises of the 80s by identifying genes that are fundamental to human diseases. For example, the genes defective in cystic fibrosis,<sup>1,2</sup> neurofibromatosis,<sup>3</sup> and colorectal cancer<sup>4-8</sup> are known or are beginning to be established. Secondly, alongside these advances a revolution has occurred in the methodology of molecular biology with the development of the polymerase chain reaction (PCR).<sup>9,10</sup>

PCR has been described by Alec Jeffries as 'the molecular biologists' answer to photocopying'. It allows the generation of millions of exact copies of specific pieces of nucleic acid and has changed the practice of molecular biology in the short period since its introduction. Molecular biologists were previously limited by the minute amounts of the gene of interest when compared with the large excess of the background DNA. The choices which faced them were either to clone and replicate the gene of interest in bacterial hosts or to use sensitive radioactive identification methods such as Southern or northern blotting. For many situations these have now been superseded by PCR based techniques which enable new approaches to these problems and produce results in hours or days rather than weeks. Importantly, it is no longer necessary to beg, borrow, or buy a range of probes before starting new work, you can synthesise the primers yourself or order them from within your institution or hospital at no greater price than a monoclonal antibody. Furthermore, as soon as an exciting advance has been reported it is possible to reproduce rapidly the experiments within your own laboratory. The transfer of knowledge no longer takes months or years but days. The cystic fibrosis gene 3 base pair deletion<sup>1,2</sup> is an excellent example, in that within weeks of its description, secondary publications were blossoming within the letter columns of the *Lancet*.<sup>11-13</sup>

How does this technique work? Firstly, the area to be amplified on the gene of interest must be studied to identify two lengths of DNA, usually 20 base pairs long, that surround this area. Complimentary sequences (primers) are then built from the individual deoxynucleotides on a DNA synthesiser. The primers are the key to the reaction as they

guarantee its specificity. The DNA is firstly denatured to single stranded DNA by heating to 95°C. The primers bind to their complementary sites when the temperature is reduced to around 55°C, forming two small areas of double stranded DNA. This acts as a template for DNA synthesis when the temperature is raised to 72°C. The DNA polymerase, usually Taq polymerase, then begins to make double stranded DNA starting at each primer site producing two copies of the site of interest. How does PCR generate millions of copies? The simple answer is to repeat the whole reaction 20-40 times obtaining an exponential accumulation of the gene of interest. The reaction product is now so rich in amplified DNA that when run on a simple agarose gel a band of DNA of identical length to the distance between the primers appears, confirming the presence and size of the gene in the original DNA. Thus, someone with cystic fibrosis would have an amplified product three base pairs shorter than a normal person because of the three base pair deletion, and an individual heterozygous for the gene would produce two separate bands.

The sensitivity of the technique is exquisite, with detection limits of a single cell,<sup>14</sup> virus, or bacterium, unlike other molecular biological techniques. Furthermore, the quality of DNA present can be very poor. This means that DNA can be amplified from most clinical specimens, paraffin embedded material,<sup>15,16</sup> and even from paleological material as old as 17 million years.<sup>17</sup>

How will PCR revolutionise gastroenterology? Four areas will initially feel the impact of this technique – microbiology, oncology, genetics, and immunology. These areas will be developed in more detail in the rest of this series, but will be briefly touched upon here.

#### Microbiology

The diagnosis of viral infection is often by serology and less often by the direct detection of the virus by time consuming or expensive tests. PCR allows the rapid (within five hours) and definitive diagnosis of the presence of a particular virus, and even its typing. The material can be from a variety of sources, and even contaminated with other organisms. Faecal detection of organisms has been made much easier,<sup>18</sup> but more work is required to identify the optimum extraction methods. PCRs to over 20 different viruses have been

developed, many with a direct gastrointestinal interest such as hepatitis A,<sup>19</sup> B,<sup>20</sup> and C,<sup>21</sup> enteroviruses;<sup>22</sup> adenoviruses;<sup>23</sup> human papilloma viruses;<sup>24</sup> measles etc.<sup>25</sup>

It is also possible to diagnose bacterial infections rapidly by the demonstration of bacterial DNA, avoiding culture and biochemical analysis. This technology has the greatest value in detecting the presence of mycobacteria such as *Mycobacterium tuberculosis*,<sup>26</sup> *M paratuberculosis*,<sup>27, 28</sup> and *M leprae*,<sup>29</sup> where it is possible to avoid six weeks or more of culture. PCR also allows the direct demonstration of organisms such as *Treponema pallidum*.<sup>30</sup> If the presence of a bacterial toxin is of more interest than the bacteria itself, then toxin carrying strains can be identified by designing primers specific to the toxin gene, as reported for *Clostridium difficile*<sup>31</sup> and *Escherichia coli*.<sup>32</sup> The presence of antibiotic resistant genes could also be excluded, allowing earlier treatment with drugs to which the organism is sensitive. This technique also allows the search for unknown organisms by the use of universal primers to 16S rRNA genes, which amplify any bacteria in the material to be studied.<sup>33</sup> This can then be sequenced and the organism identified. Specific 16S rRNA primers can also be designed as described for organisms such as *Helicobacter pylori*.<sup>34</sup>

### Oncology

In oncology the use of PCR is currently limited to the identification of translocations, monoclonality,<sup>35</sup> and of minimal residual disease<sup>36</sup> before clinical relapse in lymphomas. However, should any of the recently described molecular abnormalities of colorectal cancer (Kirsten-ras,<sup>5</sup> the DCC gene,<sup>6</sup> p53 deletion,<sup>4</sup> the APC gene<sup>7, 8</sup>) prove to have value in diagnosis or prognosis then these can be rapidly used. The recent description of the APC gene<sup>7, 8</sup> will make genetic diagnosis in this condition much easier and will advance our knowledge of the early stage of colorectal neoplasia. PCR can also be used to search for molecular markers of biological aggressiveness in archival tissue, thus removing the period of follow up required for the assessment of their impact on prognosis.

### Genetics

The diagnosis of genetic diseases will be simplified by PCR. The parental origin of chromosomes can be traced by PCR detection of inherited polymorphisms such as variable number tandem repeats, microsatellite tandem repeats,<sup>37</sup> and alu<sup>38</sup> or L1H repeat sequences, thus enabling the risk of a genetic disease to be assessed without exact knowledge as to the causative gene. If the gene has been identified, then direct confirmation can be obtained. In the future it may be possible for the gastrointestinal pathologist to go straight from the haematoxylin and eosin slide or paraffin block to an exact molecular diagnosis of a genetic condition without further sampling being needed.

### Immunology

In immunology and organ transplantation, the HLA typing of samples will be replaced by an accurate PCR diagnosis of the genetic sequence of the individual<sup>39, 40</sup> and much will be learned of the intricacies of HLA disease associations. The level of cytokine expression can also be determined.<sup>41</sup>

### Other aspects of the new technology

A range of diagnostic tests is now available, but this is not the limit of the technology. It can provide relatively large amounts of nucleic acid for direct DNA<sup>42</sup> or RNA sequencing, it can create modified nucleic acid sequences<sup>43</sup> which can be

inserted into plasmids or transgenic animals to model genetic abnormalities, and can even be used to engineer genetically antibodies for therapy or diagnostic purposes. New genes can be identified using sequences conserved within gene families such as the tyrosine kinases.<sup>44</sup> It may seem unlikely that these molecular techniques will become routine tools of the hospital laboratory but it should be remembered that DNA sequencing has been taken up by many regional DNA laboratories and today's research techniques can rapidly become tomorrow's routine tool, especially if gastroenterologists are aware of what is on offer from the molecular pathologists.

PHILIP QUIRKE

Academic Unit,  
Department of Histopathology,  
University of Leeds and  
United Leeds Hospitals  
NHS Trust,  
Leeds

- Riordan JR, Rommens JM, Kerem B, Alon N, Rozmahel R, Grzelczak Z, et al. Identification of the cystic fibrosis gene: cloning and characterisation of complementary DNA. *Science* 1989; 245: 1066-73.
- Kerem B, Rommens JM, Buchanan JA, Markiewicz D, Cox TK, Chakravarti A, et al. Identification of the cystic fibrosis gene: genetic analysis. *Science* 1989; 245: 1073-80.
- Wallace MR, Marchuk DA, Andersen LB, Letcher R, Oden HM, Saulino AM, et al. Type 1 neurofibromatosis gene: identification of a large transcript disrupted in three NF1 patients. *Science* 1990; 249: 181-6.
- Baker SJ, Fearon ER, Nigro JM, Hamilton SR, Preisinger AC, Jessup JM, et al. Chromosome 17 deletions and p53 gene mutations in colorectal carcinomas. *Science* 1989; 244: 217-21.
- Bos JL, Fearon ER, Hamilton SR, Verlaan-de Vries M, van Boom JH, van der Eb AJ, et al. Prevalence of ras gene mutations in human colorectal cancers. *Nature* 1987; 327: 293-7.
- Fearon ER, Cho KR, Nigro JM, Kern SE, Simons JW, Ruppert JM, et al. Identification of a chromosome 18q gene that is altered in colorectal cancers. *Science* 1990; 247: 49-56.
- Kinzler KW, Nilbert MC, Su L, Vogelstein B, Bryan TM, Levy DB, et al. Identification of FAP focus genes from chromosome 5q21. *Science* 1991; 253: 661-5.
- Nishisho I, Nakamura Y, Miyoshi Y, Miki Y, Ando H, Horii A, et al. Mutations of chromosome 5q21 genes in FAP and colorectal cancer patients. *Science* 1991; 253: 665-9.
- Mullis KB, Faloona FA. Specific synthesis of DNA *in vitro* via a polymerase-catalyzed chain reaction. *Methods in Enzymol* 1987; 155: 335-50.
- Saiki RK, Scharf S, Faloona F, Mullis KB, Horn GT, Erlich HA, et al. Enzymatic amplification of beta-globin genomic sequences and restriction site analysis for diagnosis of sickle cell anaemia. *Science* 1985; 230: 1350-4.
- Bowling FG, McGill JJ, Sheperd RW, Danks DM. Screening for cystic fibrosis: use of delta F508 mutation. *Lancet* 1990; i: 925-6.
- Newton CR, Schwarz M, Summers C, Heptinstall LE, Graham A, Smith JC, et al. Detection of the delta F 508 deletion by amplification refractory mutation system. *Lancet* 1990; i: 1217-9.
- Dumar V, Lafitte JJ, Gervais R, Debaeche D, Kesteloot M, Lalau G, et al. Abnormal distribution of cystic fibrosis delta F508 allele in adults with chronic bronchial hypersecretion. *Lancet* 1990; i: 1340.
- Li HH, Gyllenstein UB, Cui XF, Saiki RK, Erlich HA, Arnheim N. Amplification and analysis of DNA sequences in single human sperm and diploid cells. *Nature* 1988; 335: 414-7.
- Impraim CC, Saiki RK, Erlich HA, Teplitz RL. Analysis of DNA extracted from formalin-fixed, paraffin-embedded tissues by enzymatic amplification and hybridisation with sequence-specific oligonucleotides. *Biochem Biophys Res Commun* 1987; 142: 710-6.
- Jackson DP, Lewis FA, Taylor GR, Boylston AW, Quirke P. Tissue extraction of DNA and RNA and analysis by the polymerase chain reaction. *J Clin Pathol* 1990; 43: 499-504.
- Paabo S, Higuchi RG, Wilson AC. Ancient DNA and the polymerase chain reaction. *J Biol Chem* 1989; 264: 9709-12.
- Wilde J, Eiden J, Yolken R. Removal of inhibitory substances from human faecal specimens for detection of group A Rotaviruses by reverse transcriptase and polymerase chain reaction. *J Clin Microbiol* 1990; 28: 1300-7.
- Jansen RW, Siegl G, Lemon SM. Molecular epidemiology of human hepatitis A virus defined by an antigen-capture polymerase chain reaction method. *Proc Natl Acad Sci USA* 1990; 87(8): 2867-71.
- Lo Y-MD, Mehal WZ, Fleming KA. In vitro amplification of hepatitis B virus sequences from liver tumour DNA and from paraffin wax embedded tissues using the polymerase chain reaction. *J Clin Pathol* 1989; 42: 840-6.
- Garson JA, Tedder RS, Briggs M, Tuke P, Glazebrook JA, Trute A, et al. Detection of hepatitis C viral sequences in blood donations by 'nested' polymerase chain reaction and prediction of infectivity. *Lancet* 1990; 335: 1419-22.
- Chapman NM, Tracy S, Gauntt CJ, Fortmueller U. Molecular detection and identification of enteroviruses using enzymatic amplification and nucleic acid hybridization. *J Clin Microbiol* 1990; 28: 843-50.
- Allard A, Girones R, Juto P, Wadell G. Polymerase chain reaction for detection of adenoviruses in stool samples. *J Clin Microbiol* 1990; 28: 2659-67.
- Shimada M, Fukushima M, Mukai H, Kato I, Nishikawa A, Fujinaga K, et al. Amplification and specific detection of transforming gene region of human papillomavirus 16, 18 and 33 in cervical carcinoma by means of the polymerase chain reaction. *Jap J Cancer Res* 1990; 81: 1-5.
- Jackson DP, Quirke P, Lewis FA, Boylston AW, Sloan JM, Robertson D, et al. Detection of measles virus RNA in paraffin embedded tissue. *Lancet* 1989; i: 1391.
- Brisson-Noël A, Aznar C, Churcau C, Nguyen S, Pierre C, Bartoli M, et al. Diagnosis of tuberculosis by DNA amplification in clinical practice evaluation. *Lancet* 1991; 338: 364-6.

- 27 Quirke P, Dockey D, Taylor GR, Lewis FA, Hawkey P, Graham D, *et al.* Detection of *Mycobacterium paratuberculosis* in inflammatory bowel disease. *Gut* 1991; **32**: A572.
- 28 Sanderson JD, Moss M, Malik Z, Tizard M, Green EP, Hermon-Taylor J. Polymerase chain reaction detects *Mycobacterium paratuberculosis* in Crohn's disease tissue extracts. *Gut* 1991; **32**: A572.
- 29 Plikaytis BB, Gelber RH, Shinnick TM. Rapid and sensitive detection of mycobacterium leprae using a nested-primer gene amplification assay. *J Clin Microbiol* 1990; **28**: 1913–7.
- 30 Burstain JM, Grimprel E, Lukehart SA, Norgard MV, Radolf JD. Sensitive detection of treponema pallidum by using the polymerase chain reaction. *J Clin Microbiol* 1991; **29**: 62–9.
- 31 Kato N, Ou CY, Kato H, Bartley SL, Brown VK, Dowel VR, *et al.* Identification of toxigenic clostridium difficile by the polymerase chain reaction. *J Clin Microbiol* 1991; **29**: 33–7.
- 32 Olive M. Detection of enterotoxigenic *Escherichia coli* after polymerase chain reaction amplification with a thermostable DNA polymerase. *J Clin Microbiol* 1989; **27**(2): 261–5.
- 33 Ward DM, Weller R, Bateson MM. 16S rRNA sequences reveal numerous uncultured microorganisms in a natural community. *Nature* 1990; **345**: 63–5.
- 34 Ho SA, Lewis F, Wyatt JI, Dixon MF, Secher DA, Tompkins DS, *et al.* *Helicobacter pylori* detection by PCR of the gene encoding 16S ribosomal RNA in fresh and paraffin embedded tissue. *J Pathol* 1990; **161**: 351(A).
- 35 Wan JH, Trinor KJ, Brisco MJ, Morley AA. Monoclonality in B cell lymphoma detected in paraffin wax embedded sections using the polymerase chain reaction. *J Clin Pathol* 1990; **43**: 888–90.
- 36 Ugozzoli L, Yam P, Petz LD, Ferrara GB, Champlin RE, Forman SJ, *et al.* Amplification by the polymerase chain reaction of hypervariable regions of the human genome for the evaluation of chimerism after bone marrow transplantation. *Blood* 1991; **77**: 1607–15.
- 37 Weber JL, May PE. Abundant Class of Human DNA polymorphisms which can be typed using the polymerase chain reaction. *Am J Hum Genet* 1989; **44**: 388–96.
- 38 Britten RJ, Baron WF, Stout DB, Davidson EH. Sources and evolution of human Aeu repeated sequences. *Proc Natl Acad Sci USA* 1988; **85**: 4770–4.
- 39 Gyllensten UB, Erlich HA. Generation of single-stranded DNA by the polymerase chain reaction and its application to direct sequencing of the HLA-DQA locus. *Proc Natl Acad Sci USA* 1988; **85**: 7652–6.
- 40 Bell J. The polymerase chain reaction. *Immunol Today* 1989; **10**(10): 351–5.
- 41 Gilliland G, Perrin S, Blanchard K, Bunn HF. Analysis of cytokine mRNA and DNA: detection and quantitation by competitive polymerase chain reaction. *Proc Natl Acad Sci USA* 1990; **87**(7): 2725–9.
- 42 Engelke DR, Hoener PA, Collins FS. Direct sequencing of enzymatically amplified human genomic DNA. *Proc Natl Acad Sci USA* 1988; **85**: 544–8.
- 43 Clackson T, Güssow D, Jones PT. General applications of PCR to gene cloning and manipulations. In: McPherson MJ, Quirke P, Taylor GR, eds. *PCR: A practical approach*. Oxford: IRL Press, 1991: 187–214.
- 44 Wilks AF. Two putative protein-tyrosine kinases identified by application of the polymerase chain reaction. *Proc Natl Acad Sci USA* 1989; **86**: 1603–7.