

Blood and brain concentrations of mercaptans in hepatic and methanethiol induced coma

H AL MARDINI, K BARTLETT, AND C O RECORD

From the Gastroenterology Unit and Department of Clinical Biochemistry and Metabolic Medicine, Royal Victoria Infirmary and University of Newcastle upon Tyne

SUMMARY It has been suggested that mercaptans are important factors in the pathogenesis of hepatic encephalopathy. Using a gas liquid chromatographic technique which uses propanethiol as internal standard and a sulphur specific detector, blood methanethiol concentration was found to be significantly risen in encephalopathic patients with liver disease (13.2 ± 1.0 nmol/ml; $n=47$) compared with control subjects (5.7 ± 0.3 , $n=29$; $p<0.05$) and non-encephalopathic patients with liver disease (7.7 ± 1.1 , $n=35$; $p<0.05$) but ethanethiol and dimethylsulphide concentrations were similar in the three groups. Blood methanethiol, however, was not clearly related to coma grade, similar values being found in deeply comatosed patients to those showing only mild cerebral dysfunction. When rats were injected with 40–120 μ mol methanethiol a range of responses was obtained which varied between normal consciousness and coma. The minimum blood concentration of methanethiol associated with coma (200 nmol/ml) was at least 10-fold greater than in patients with hepatic encephalopathy but brain concentrations were similar in comatose rats and those which remained awake. Blood methanethiol concentrations were similar in control and germ free rats and did not rise in cirrhotics or controls after ingestion of 2 g methionine. It is concluded that while methanethiol may accumulate in hepatic coma, it is unlikely to be of major pathogenetic importance. Endogenous mercaptans are unlikely to originate from bacterial metabolism in the gut.

The precise pathogenesis of hepatic encephalopathy is poorly understood although a number of substances have been implicated because they cause coma in experimental animals. These include ammonia, short chain fatty acids and mercaptans and synergism between these substances has been shown.¹ Other possible mechanisms include an accumulation of false neurotransmitters such as octopamine and a fall in normal neurotransmitters such as dopamine and noradrenaline.² Recent studies in animals, however, have shown a marked fall in cerebral neurotransmitters after intraventricular injection of octopamine while no apparent change in consciousness was observed.³ Furthermore noradrenaline concentrations were similar in the brains of patients with hepatic encephalopathy compared with controls while

dopamine concentrations were normal or increased.⁴ Brain octopamine concentrations were not significantly greater than in controls.⁴

Mercaptans were first implicated in hepatic coma when methanethiol was isolated from the urine.⁵ Subsequent investigations showed increased concentrations of methanethiol⁶ and dimethylsulphide⁷ in the breath of patients with hepatic failure. In a recent study methanethiol was significantly increased in the blood of patients with hepatic encephalopathy, 93% of encephalopathic patients having values outside the normal range.⁸ Ethanethiol also appears to be significantly increased.⁹ In the present study we have assessed concentrations in blood of the mercaptans methanethiol, ethanethiol, and dimethylsulphide in patients with different grades of hepatic coma and compared them with values in rats in coma after an intraperitoneal injection of methanethiol. We have also compared brain concentrations of mercaptans in rats with experimental liver damage with methanethiol induced coma. As the origin of blood mercaptans is

Address for correspondence: Dr C O Record, Gastroenterology Unit, The Royal Victoria Infirmary, Queen Victoria Road, Newcastle upon Tyne NE1 4LP.

Received for publication 31 May 1983

at present unclear we have also assessed changes in blood concentrations in patients with cirrhosis after ingestion of methionine and examined mercaptan concentrations in germ free rats.

Methods

PATIENTS

Blood was obtained by standard venepuncture from patients with hepatic encephalopathy, patients with liver disease but no evidence of encephalopathy, and control subjects. All decompensated patients with hepatic encephalopathy were studied irrespective of the aetiology of the liver disease or whether acute or chronic. Four 1 ml aliquots of whole blood were added to ice cold 7 ml septum bottles and immediately frozen before assay by gas liquid chromatography as previously described.¹⁰ The frozen blood samples in closed septum bottles were thawed, 200 μ l propanethiol internal standard solution (100 μ l propanethiol in 5 ml methanol diluted to 42 μ mol/l with water) added to each sample *via* a slide valve and septum (Minnert), shaken for 30 seconds and refrozen. The blood samples were then treated with zinc and phosphoric acid to obtain maximal release of mercaptans. For this 1 ml zinc slurry (10 g zinc dust stirred in 50 ml water at 55°C) and 0.5 ml of 3.3 mol/l phosphoric acid were added to the sample and heated at 60°C for 10 minutes. To sample the gas phase 1 ml air was injected into the sample bottle and 1 ml withdrawn and injected onto the gas chromatograph. Gas samples were analysed using a Carlo Erba model FV 350 gas chromatograph fitted with sulphur specific detector. A glass column (2.5 mm \times 3 mm internal diameter) packed with 20% SE30 on 60–80 Chromosorb P was used. The gas flow rates were as follows: nitrogen carrier gas 130 ml/min, hydrogen 45 ml/min, oxygen 12.5 ml/min, air 15 ml/min. The column was run isothermally at 36°C and the injection port and detectors maintained at 100°C. Samples were injected onto the column with a 1 ml A2 pressure lock gas syringe (Precision Sampling Corporation, USA). The chromatograph was fitted with both a pen recorder and integrator (Infotronics, UK Model 308).

Encephalopathic patients were graded for depth of coma, using the classification of Trey and Davidson.¹¹ Where possible sequential estimations were undertaken. To investigate the origin of mercaptans in blood, five fasting outpatients with stable biopsy diagnosed cirrhosis gave informed consent to ingest 2 g methionine in tablet form. Blood was obtained at hourly intervals over the next four hours. For estimation of blood mercaptan concentration in germ free rats, blood was obtained

by decapitation. Brain mercaptan estimations were undertaken on 200 g rats treated with a lethal dose of carbon tetrachloride (3.2 g/kg). The rats were killed 24 hours after carbon tetrachloride administration, when liver function tests were severely deranged or sooner if they appeared moribund. The majority of rats treated with this dose of carbon tetrachloride died within the following 48 hours. The cortex was placed on ice, weighed and added to four parts Tris HCl buffer (pH 7.0) before sonication. One millilitre aliquots were subsequently added to septum bottles and treated in the same way as whole blood.

Methanethiol induced coma was induced by intraperitoneal injection of a solution of methanethiol in water. The latter was prepared by passing a stream of methanethiol gas (Cambrian Gases, Croydon, Surrey, UK) through water as previously described.¹⁰ The dose of methanethiol administered varied between 40 and 120 μ mol.

STATISTICAL METHODS

Values quoted are mean \pm SEM. P values refer to the Student's *t* test or the Mann-Whitney U test where appropriate.

Results

BLOOD MERCAPTANS IN PATIENTS FOLLOWING INGESTION OF 2 G METHIONINE

Concentrations of blood methanethiol (Fig. 1), ethanethiol and dimethylsulphide (Fig. 2) after ingestion of 2 g methionine are shown. There were no significant differences over the observation period.

BLOOD AND BRAIN MERCAPTANS IN GERM FREE RATS

Blood methanethiol, ethanethiol, and dimethylsulphide concentrations were similar in germ free

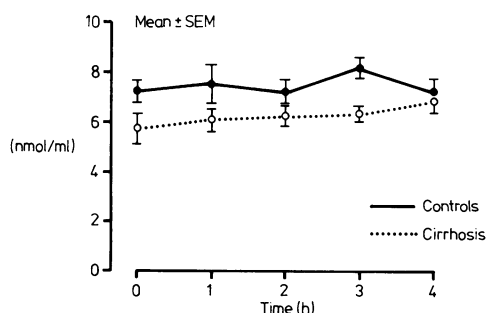
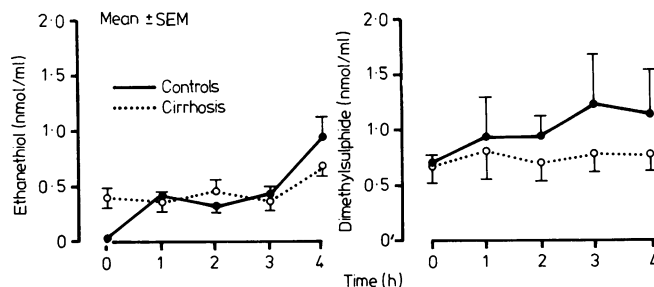


Fig. 1 Blood methanethiol after ingestion of 2 g of methionine by patients with cirrhosis and controls.

Fig. 2 Blood ethanethiol and dimethylsulphide after ingestion of 2 g of methionine by patients with cirrhosis and controls.



and control rats, no significant differences being observed (Table 1). Brain methanethiol and ethanethiol concentrations were also similar, but dimethylsulphide concentration was significantly increased in germ free rats compared with controls ($p < 0.01$; Table 1).

BLOOD MERCAPTANS IN PATIENTS WITH LIVER DISEASE

Blood mercaptan concentrations in patients with liver disease were divided into those exhibiting hepatic encephalopathy (31) and non-encephalopathic control subjects (35). Results were compared with 29 normal subjects. Methanethiol was significantly increased in patients with encephalopathy (47 observations) compared with those without encephalopathy and control subjects ($p < 0.05$), but ethanethiol and dimethylsulphide concentrations were similar in the three groups (Table 2). When patients were further subdivided according to their grade of coma (Fig. 3) there was a stepwise increase in mean concentration of methanethiol with increasing coma grade. There was considerable overlap, however, between groups, the highest value being seen in a non-encephalopathic patient. The differences between the different grades of coma were not significant.

BRAIN CONCENTRATIONS OF MERCAPTANS

Brain concentrations of methanethiol, ethanethiol,

and dimethylsulphide in rats treated with carbon tetrachloride are shown in Table 3. Blood concentrations (previously reported¹⁰) are also shown. Brain concentrations were similar to controls. For methanethiol simultaneous blood concentrations were significantly increased but this did not lead to a rise in brain concentration.

METHANETHIOL INDUCED COMA

Concentrations of mercaptans in blood and brain in rats treated with intraperitoneal methanethiol (40–120 μmol) are shown in Figures 4 and 5. Rats were divided into those remaining awake, those just unable to stand but not comatose, and those in coma within three minutes of administration. When rats in coma were left they regained consciousness over the succeeding 15 minutes. Rats were killed three minutes after methanethiol if still awake or sooner if comatose.

Blood methanethiol concentrations in rats remaining awake were considerably greater than in patients with hepatic coma (Fig. 4). Rats just unable to stand and those in coma showed even higher concentrations so that the threshold for coma appeared to be associated with a blood concentration about 10-fold greater than in patients with hepatic coma. There was considerable overlap in the brain concentrations of methanethiol, some rats in coma having concentrations similar to those seen in rats which were awake (Fig. 4).

Dimethylsulphide concentrations in blood in rats

Table 1 Concentrations of mercaptans in blood and brain of germ free and control rats (mean \pm SEM)

	Blood ($\mu\text{mol/l}$)			Brain (nmol/g wet weight)		
	MT	ET	DMS	MT	ET	DMS
Germ free rats (6)	9.2 \pm 1.1	0.3 \pm 0.1	0.9 \pm 0.4	7.9 \pm 0.8	1.6 \pm 0.1	1.8 \pm 0.2
Controls (8)	7.0 \pm 0.3	0.1 \pm 0.05	1.1 \pm 0.4	7.3 \pm 0.6	1.0 \pm 0.3	0.8 \pm 0.2*

* $p < 0.01$.

MT = methanethiol; ET = ethanethiol; DMS = dimethylsulphide.

Table 2 Blood mercaptans in normal subjects and patients with liver disease ($\mu\text{mol/l}$; mean \pm SEM)

	MT	ET	DMS
Patients with liver disease			
Without			
encephalopathy (35)	7.7 \pm 1.1	0.19 \pm 0.05	0.57 \pm 0.09
With encephalopathy (47)	13.2 \pm 1.0*	0.35 \pm 0.05	0.62 \pm 0.06
Normal subjects (29)	5.7 \pm 0.3	0.30 \pm 0.05	0.42 \pm 0.04

* $p < 0.05$.

treated with methanethiol were considerably increased (Fig. 5), suggesting rapid conversion of methanethiol *in vivo* to dimethylsulphide. Brain concentrations were also increased, but concentrations in rats which were awake overlapped with those seen in comatose rats (Fig. 5).

BLOOD AMMONIA CONCENTRATIONS

Simultaneous blood ammonia and methanethiol concentrations in patients with hepatic coma are shown in Figure 6. There was no correlation between ammonia concentrations and methanethiol ($r = 0.164$).

In nine patients sequential observations during different grades of coma showed that methanethiol concentrations were always higher in the more severe grade of coma (Fig. 7). These differences, however, were small and were of no value in predicting the coma grade.

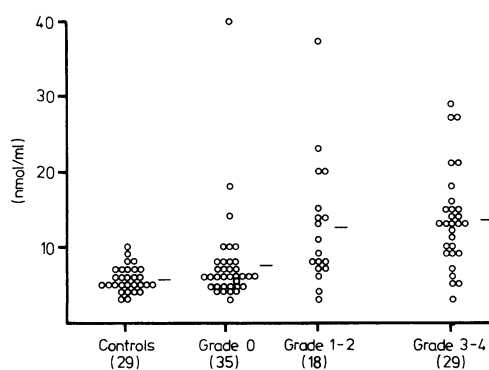


Fig. 3 Blood methanethiol concentrations in control subjects and patients with different grades of hepatic encephalopathy. Mean values are shown by solid bars with number of observations in parenthesis. There were no significant differences between any groups. Coma was graded as described by Trey and Davidson.¹¹

Discussion

In the present study we have extended the previous observations of Doizaki and Zieve¹² and McClain⁸ to include determinations of ethanethiol and dimethylsulphide in the blood and have also measured all three mercaptans in the brain. Unfortunately, these earlier studies were marred by the variable recovery of methanethiol from blood which ranged from 25–80%, higher recoveries being found with the higher blood concentrations. The present studies have used an improved gas chromatographic method in which recovery was constant over a wide concentration range.¹⁰ Using this method blood methanethiol concentrations in normal man were 10-fold higher than that reported by McClain⁸ but were 10-fold lower than that found by Brunner and Scharff.¹³ Because of the wide discrepancy between our normal range and that found by others we have determined the minimum blood concentration of methanethiol which was associated with coma after administration to rats. By injecting between 40 and 120 μmol of methanethiol a range of responses was obtained which varied between normal consciousness, just able to maintain upright posture, and coma. The minimum blood concentration of methanethiol which was associated with coma was 200 $\mu\text{mol/l}$ while the mean concentration in patients with hepatic encephalopathy was 14.5 $\mu\text{mol/l}$, the highest value observed in patients with liver disease being 40 $\mu\text{mol/l}$. Zieve *et al*¹ have suggested that methanethiol, ammonia, and short chain fatty acids act synergistically and that subcoma doses of each given individually are able to induce coma when given together. When subcoma doses of methanethiol were given with ammonia or short chain fatty acids and coma resulted, however, blood concentrations of methanethiol were similar to those in rats given sufficient methanethiol alone to produce coma. It has previously been suggested that rats with such artificial coma exhibited similar concentrations of methanethiol in blood to rats and man with hepatic encephalopathy^{8 12} but the present findings do not support this view, a 10-fold difference being found. Acute changes in the rat following methanethiol administration may not be comparable to the more slowly evolving hepatic coma in man. This factor, however, is unlikely to account for the large differences in blood methanethiol concentrations associated with coma in the two groups.

Previous studies⁸ have suggested that methanethiol determinations are valuable in patients with hepatic coma. In the present study we have found considerable overlap in the blood concentrations of methanethiol in patients with different grades of coma and control subjects. Indeed the highest

Table 3 Concentrations of mercaptans in blood and brain of rats treated with carbon tetrachloride (3.2 g/kg) and controls

	Blood			Brain		
	MT	ET	DMS	MT	ET	DMS
Carbon tetrachloride	12.5±0.8* (14)	0.1±0.1 (14)	1.6±0.4 (14)	7.8±0.4 (12)	1.4±0.2 (12)	1.1±0.1 (12)
Controls	7.0±0.3 (9)	0.1±0.05 (9)	1.1±0.4 (9)	7.3±0.6 (8)	1.0±0.3 (8)	0.8±0.2 (8)

Concentrations of mercaptans are expressed as $\mu\text{mol/l}$ for blood and nmol/g wet weight for brain, as mean \pm SEM with number observations in parenthesis. Values for carbon tetrachloride treated rats which are statistically different from controls are indicated by * $p < 0.001$.

Rats were killed 24 h after carbon tetrachloride administration or sooner if moribund.

Fig. 4 Methanethiol concentrations in blood and brain of rats after intraperitoneal injection. Rats were divided into those which were fully awake, those just able to stand, and those which were comatose.

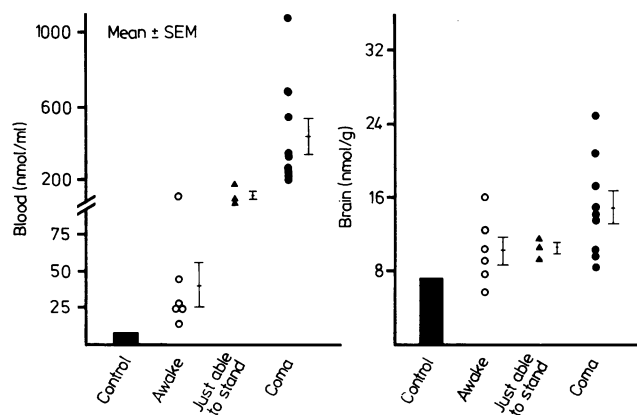
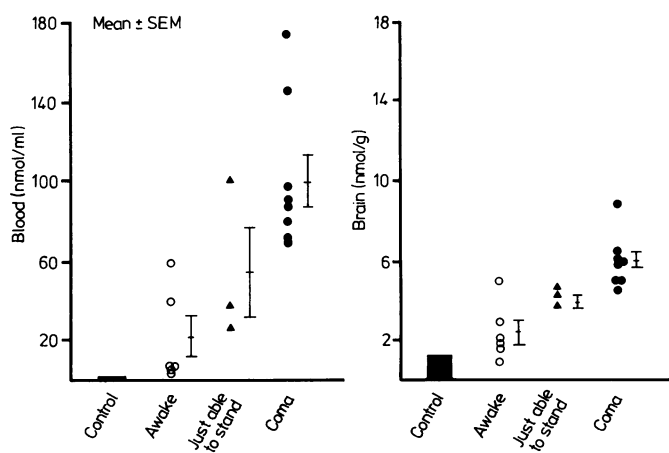


Fig. 5 Dimethylsulphide concentrations in blood and brain of rats after intraperitoneal injection of methanethiol. Rats were divided into those which were awake, those just able to stand, and those in coma.



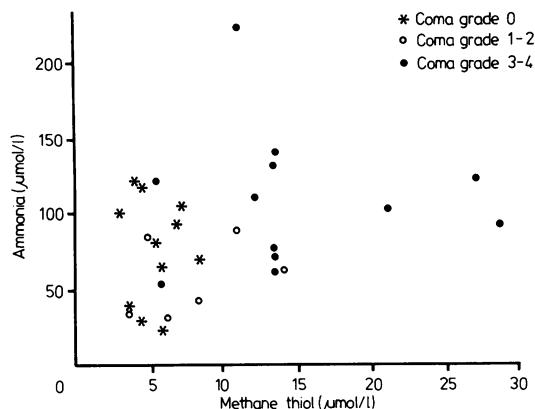


Fig. 6 Relationship between ammonia and methanethiol concentrations in patients with hepatic encephalopathy. Correlation was not significant ($r=0.164$). The coma was graded as described by Trey and Davidson.¹¹

concentration was found in a non-comatose outpatient with primary biliary cirrhosis. The mean concentration of methanethiol in encephalopathic patients, however, was significantly increased compared to non-encephalopathic patients and controls but ethanethiol and dimethylsulphide concentrations were similar in all groups. Blood ethanethiol concentrations were similar to those previously associated with coma in the rat (0.2 µmol/l)¹ while blood dimethylsulphide concentrations were very much lower than those associated with coma in the rat (7 mmol/l).¹ Valid comparisons between these figures cannot be made because of methodological differences. Both in the present

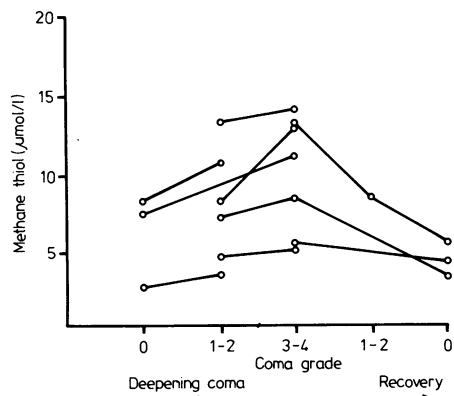


Fig. 7 Sequential methanethiol concentrations in nine patients with hepatic encephalopathy. Encephalopathy was graded as described by Trey and Davidson.¹¹

studies and those of McClain⁸ there was no correlation between blood ammonia and methanethiol concentrations.

Phear *et al*¹⁴ showed neuropsychiatric changes in cirrhotics fed methionine, an effect which occurred without change in blood ammonia concentrations and which could be prevented by the administration of chlortetracycline. Patients developing encephalopathy received a mean of 27 g methionine over two and a half days. It was suggested that this effect of methionine was due to a toxic breakdown product other than ammonia. In the present study blood methanethiol, ethanethiol, and dimethylsulphide did not change significantly after ingestion of 2 g of methionine and decreased concentrations were not observed in germ free rats. It thus seems unlikely that bacterial metabolism in the gut is the major source of endogenous mercaptans in man. Whether methanethiol was the toxic metabolite of methionine postulated by Phear¹⁴ cannot be determined from the present study because of the large difference in methionine dose administered.

Brain concentrations in rats in coma were significantly greater than in control rats but there was considerable overlap between rats treated with methanethiol which remained awake and those which went into coma. This suggests coma after methanethiol is not induced by this substance alone but that other factors are necessary. Whether methanethiol blocks neurotransmission or induces a metabolic modification in the brain is unknown.

Dimethylsulphide concentrations rose sharply after methanethiol administration suggesting endogenous conversion to dimethylsulphide in the circulation. This could account for the rise in dimethylsulphide concentrations in breath which has been reported in patients with cirrhosis.⁷

The present studies do not suggest that mercaptans play an important role in the pathogenesis of hepatic encephalopathy. Concentrations of methanethiol are certainly increased in the blood of many patients with hepatic coma but this appears to be the result of the severe derangement of metabolic function which occurs in patients with liver disease.

References

- 1 Zieve L, Doizaki WM, Zieve FJ. Synergism between mercaptans and ammonia or fatty acids in the production of coma: a possible role for mercaptans in the pathogenesis of hepatic coma. *J Lab Clin Med* 1974; **83**: 16-27.
- 2 Dodsworth, JM, James JH, Cummings MC, Fischer

- JE. Depletion of brain norepinephrine in acute hepatic coma. *Surgery* 1974; **75**: 811–20.
- 3 Zieve L, Olsen RL. Can hepatic coma be caused by a reduction of brain noradrenaline or dopamine? *Gut* 1977; **18**: 688–91.
 - 4 Cuilleret G, Pomier-Layrargues G, Pons F, Cadilhac J, Michel H. Changes in brain catecholamine levels in human cirrhotic hepatic encephalopathy. *Gut* 1980; **21**: 565–9.
 - 5 Challenger F, Walshe JM. Methyl mercaptans in relation to foetor hepaticus. *Biochem J* 1955; **59**: 372–5.
 - 6 Chen S, Zieve L, Mahadevan V. Mercaptans and dimethylsulphide in the breath of patients with cirrhosis of the liver. *J Lab Clin Med* 1970; **75**: 628–35.
 - 7 Kaji H, Hisamura M, Saito N, Murao M. Evaluation of volatile sulfur compounds in the expired alveolar gas in patients with liver cirrhosis. *Clin Chem Acta* 1978; **85**: 279–84.
 - 8 McClain CJ, Zieve L, Doizaki WM, Gilbertstadt S, Onstad GR. Blood methanethiol in alcoholic liver disease with and without hepatic encephalopathy. *Gut* 1980; **21**: 318–23.
 - 9 Holloway CJ, Brunner G, Schmidt E, Schmidt FW. Thiols and hepatic coma. *Art Organs* 1979; **3**: 15–9.
 - 10 Al Mardini H, Bartlett K, Record CO. An improved gas chromatographic method for the detection and quantitation of mercaptans in blood. *Clin Chim Acta* 1981; **113**: 35–41.
 - 11 Trey C, Davidson CS. The management of fulminant hepatic failure. In: Popper H, Schaffner E, eds. *Progress in liver disease*. New York: Grune and Stratton, 1970, vol 3: 282–98.
 - 12 Doizaki WM, Zieve L. An improved method for measuring blood mercaptans. *J Lab Clin Med* 1977; **90**: 849–55.
 - 13 Brunner G, Scharff P. Untersuchungen über den diagnostischen Wert der Bestimmung von Mercaptanen im Serum bei Lebererkrankungen. *Dtsch Med Wochenschr* 1978; **103**: 1796–1800.
 - 14 Phear EA, Ruebner B, Sherlock S, Summerskill WHJ. Methionine toxicity in liver disease and its prevention by chlortetracycline. *Clin Sci* 1956; **15**: 93–117.